Cut back linkers (*Eco*R I Restriction Endonuclease)
Size Fractionate (Sucrose Gradient)
Ligate to lambda vector (Lambda ZAP® II and gt11)
Package (in vitro lambda packaging extract)
Plate on *E. coli* host and amplify

Please substitute the following rewritten paragraph beginning at page 22 line 4:

(Amended) 8. Results:

gt11:	1.7×10^{11}/ml
ZAP® II	2.0×10^{10}/ml

Please substitute the following rewritten paragraph beginning at page 9 line 15:

(Amended) As representative examples of expression vectors which may be used there may be mentioned viral particles, baculovirus, phage, plasmids, phagemids, cosmids, phosmids, bacterial artificial chromosomes, viral DNA (*e.g.* vaccinia, adenovirus, foul pox virus, pseudorabies and derivatives of SV40), P1-based artificial chromosomes, yeasts plasmids, yeast artificial chromosomes and any other vectors specific for specific hosts of interest (such as bacillus, aspergillus, yeast, *etc.*) Thus, for example, the DNA may be included in any one of a variety of expression vectors for expressing a polypeptide. Such vectors include chromosomal, nonchromosomal and synthetic DNA sequences. Large numbers of suitable vectors are known to those of skill in the art, and are commercially available. The following vectors are provided by way of example; Bacterial QE70, pQE60, pQE-9 (Qiagen), psiX174, pBLUESCRIPT® SK, pBLUESCRIPT® KS (Stratagene); pTRC99a, PKK223-3, pDR540, pRIT2T (Pharmacia); Eukaryotic: pWLNEO, pXT1, pSG5 (Stratagene) pSVK3, pBPV, pMSG, pSVLSV540 (Pharmacia). Any other plasmid or vector may be used as long as they are replicable and viable in the host.

Please substitute the following rewritten paragraph beginning at page 18, final line:

(Amended) 4. Efficiency results:

gt11: 1.7 x 10^4 recombinants with 95% background

ZAP® II: 4.2 x 10^4 recombinants with 66% background

Please substitute the following rewritten paragraph beginning at page 22, line 11:

(Amended) Plates of the library prepared as described in Example 1 are used to multiply inoculate a single plate containing 200 µL of LB Amp/Meth, glycerol in each well. This step is performed using the High Density Replicating Tool (HDRT) of the Beckman BIOMEK® with a 1% bleach, water, isopropanol, air-dry sterilization cycle between each inoculation. The single plate is grown for 2h at 37°C and is then used to inoculate two white 96-well ® microtiter daughter plates containing 250 µL of LB Amp/Meth, glycerol in each well. The original single plate is incubated at 37°C for 18h, then store at -80°C. The two condensed daughter plates are incubated at 37° also for 18 h. The condensed daughter plates are then heated at 70°C for 45 min. to kill the cells and inactivate the host *E. coli* enzymes. A stock solution of 5mg/mL morphourea phenylalanyl-7-amino-4-trifluoromethyl coumarin (MuPheAFC, the 'substrate') in DMSO is diluted to 600 µM with 50 mM pH 7.5 Hepes buffer containing 0.6 mg/mL of the detergent dodecyl maltoside.

Please substitute the following two rewritten paragraphs, beginning on page 23, at line 1:

(Amended) Fifty µL of the 600 µM MuPheAFC solution is added to each of the wells of the white condensed plates with one 100 µL mix cycle using the BIOMEK® to yield a final concentration of substrate of ~ 100 µM. The fluorescence values are recorded (excitation = 400 nm, emission = 505nm) on a plate reading fluorometer immediately after addition of the substrate (t=0). The plate is incubated at 70°C for 100 min, then allowed to cool to ambient temperature for 15 additional minutes. The fluorescence values are recorded again (t=1). The values at t=0 are subtracted from the values at t=100 to determine if an active clone is present.

Rule E11 C8 (Amended) The data will indicate whether one of the clones in a particular well is hydrolyzing the substrate. In order to determine the individual clone which carries the activity, the source library plates are thawed and the individual clones are used to singly inoculate a new plate containing LB Amp'/Meth, =glyceol. As above, the plate is incubated at 37° to grow the cells, heated at 70°C to inactivate the host enzymes, and 50 μL of 600 μM MuPheAFC is added using the BIOMEK®.

Please substitute the following paragraph, beginning on page 26, at line 6:

C8 Rule E12 (Amended) Run 5 microliters on a 1% agarose gel to check the reaction.
Purify on a QIAQUICK® column (Quiagen).
Resuspend in 50 microliters H_2O.

Please substitute the following paragraph, beginning on page 23, at line 16:

C9 Rule E13 (Amended) Cut for 2 hours at 37° C.
Purify on a QIAQUICK® column (Quiagen).
Elute with 35 microliters H_2O.

In the Claims

Claims 17-40 have been cancelled without prejudice.

The following new claims have been added: